

# SIZE EXCLUSION CHROMATOGRAPHY

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# INTRODUCTION

- Size-exclusion chromatography (SEC), also called gel-filtration or gel-permeation chromatography (GPC), uses porous particles to separate molecules of different sizes.
- It is generally used to separate biological molecules, and to determine molecular weights and molecular weight distributions of polymers
- It is usually applied to large molecules or macromolecular complexes such as proteins and industrial polymers.

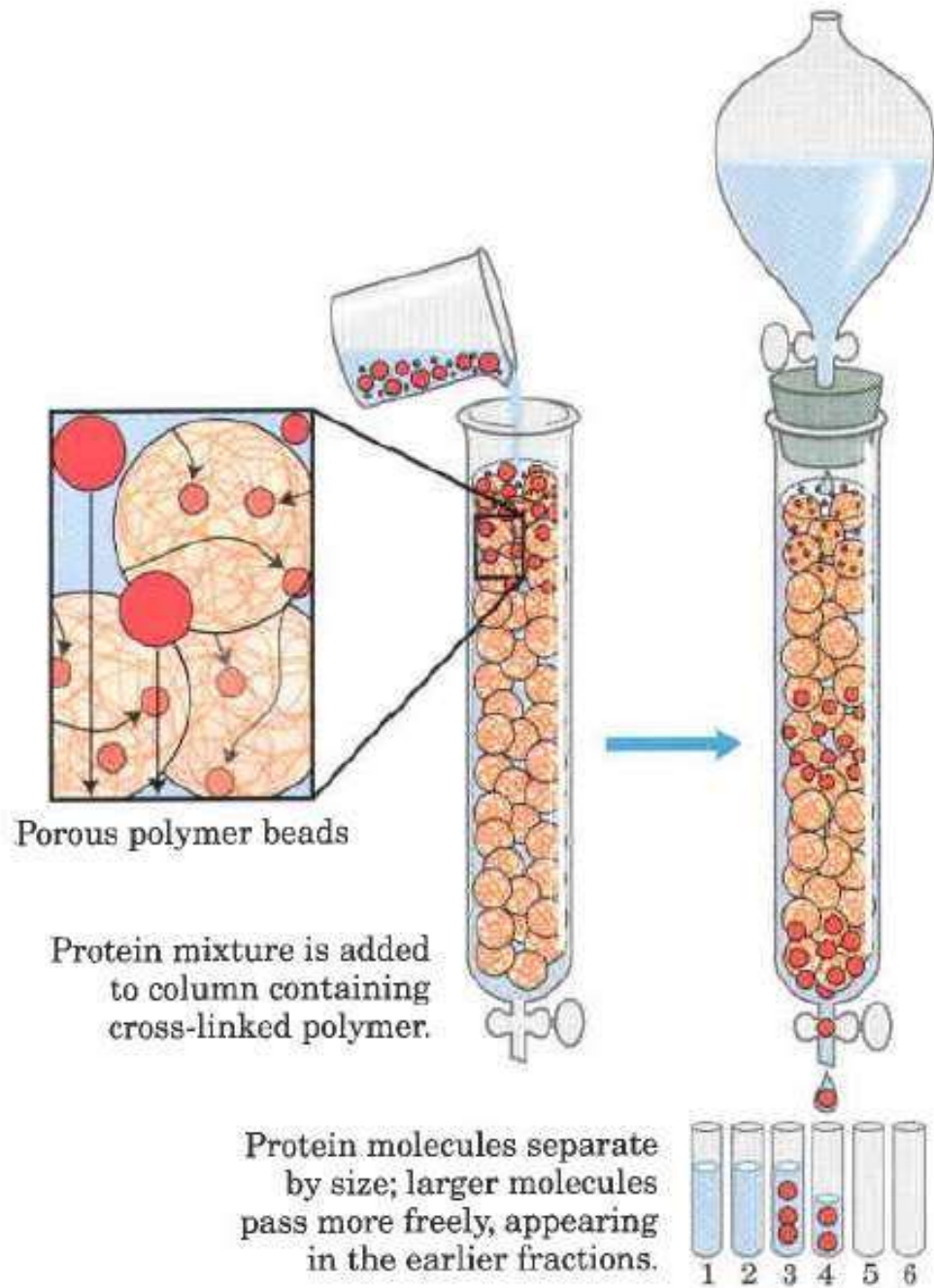
- When an aqueous solution is used to transport the sample through the column, the technique is known as Gel-filtration chromatography .
- When an organic solvent is used as a mobile phase, the technique is known as Gel-permeation chromatography.
- The separation of molecules is called **fractionation**.
- Size of pores in beads determines the **exclusion limit** (what goes through the beads and what goes around the beads)



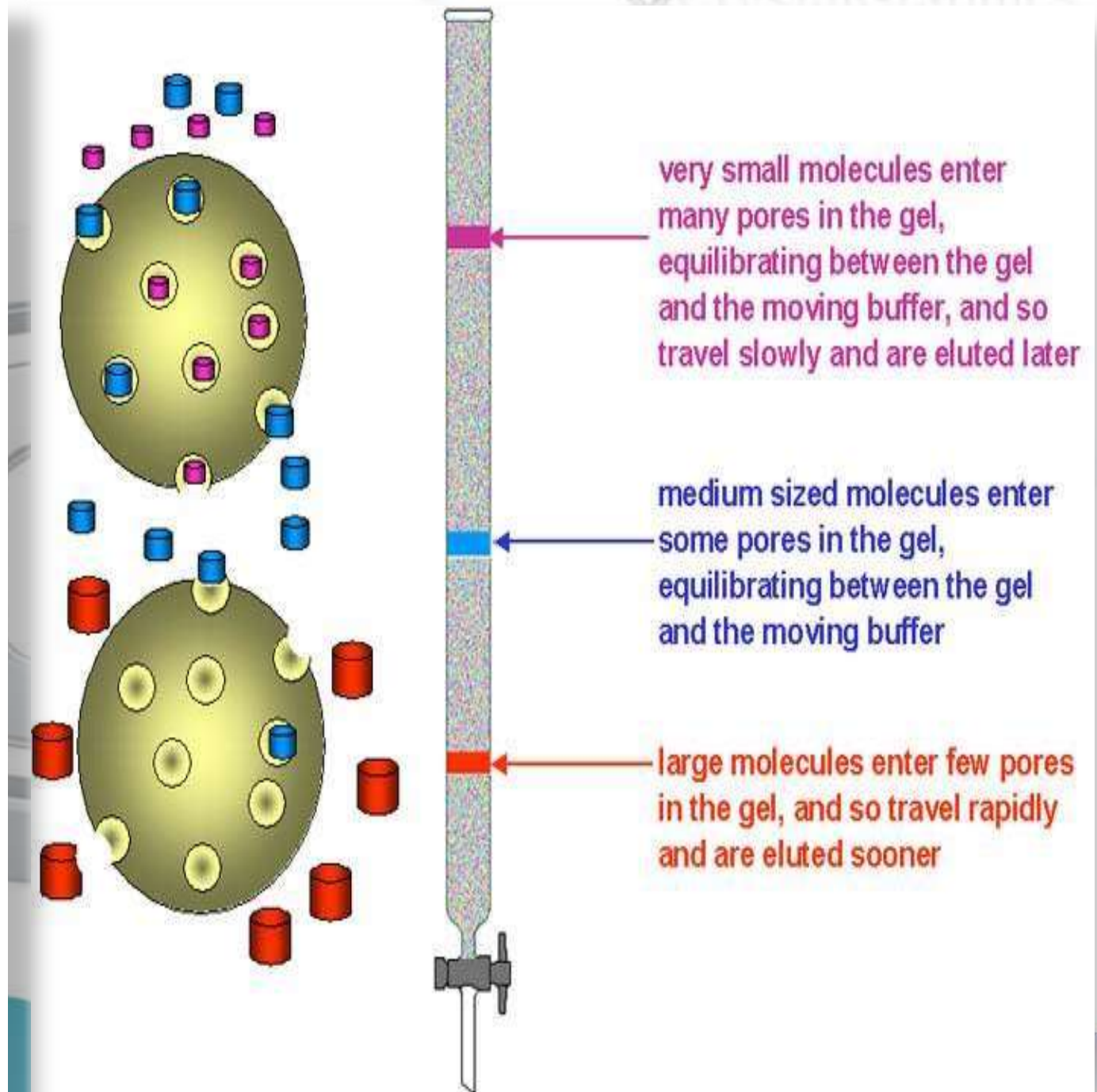
# PRINCIPLE

- A mixture of molecules dissolved in liquid (the mobile phase) is applied to a chromatography column which contains a solid support in the form of microscopic spheres, or “beads” (the stationary phase).
- The mass of beads within the column is often referred to as the column bed.
- The beads act as “traps” or “sieves” and function to filter small molecules which become temporarily trapped within the pores.

- Larger molecules pass around or are “excluded” from the beads .
- Large sample molecules cannot or can only partially penetrate the pores, whereas smaller molecules can access most or all pores.
- Thus, large molecules elute first, smaller molecules elute later, while molecules that can access all the pores elute last from the column.
- Particles of different sizes will elute (filter) through a stationary phase at different rates.



**(b) Size-exclusion chromatography**





❖ Total column volume ( $V_t$ )





$$V_t = V_g + V_i + V_0$$

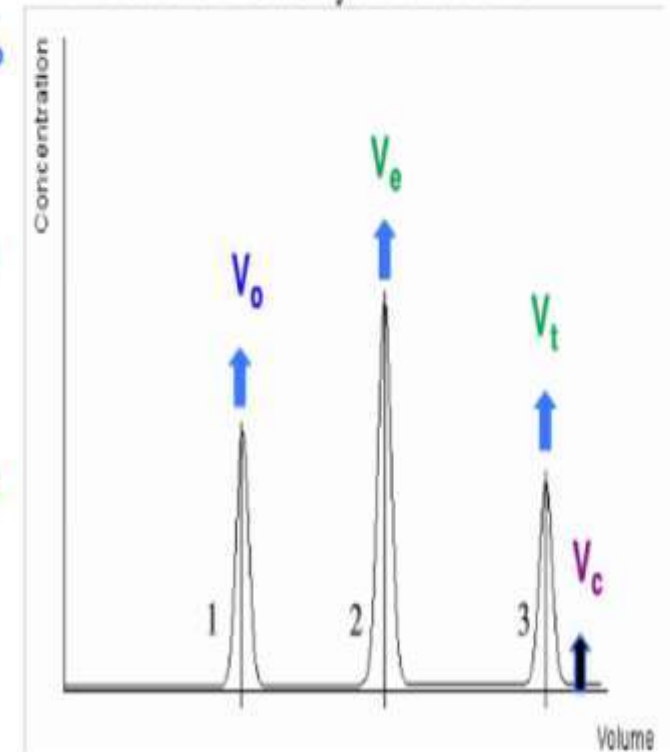
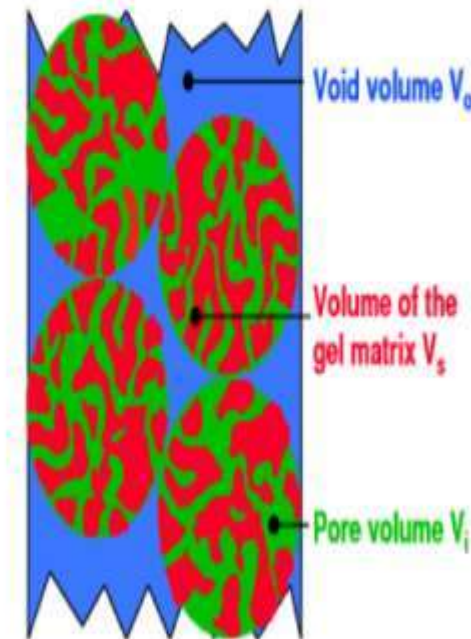
where

$V_g$  --is the volume occupied by the packing

$V_i$  --is the volume of solvent in the pores

$V_0$  --is the free solvent volume (similar to injection volume)

-   $V_0$  = Void volume: volume of the solution outside the beads, or elution from very large molecules
-   $V_e$  = the volume from the time the protein is placed until it appears in the effluent
-   $V_i$  = volume of the solution inside the beads =  $V_c - V_s - V_0$
-   $V_c$  = Total (geometric) volume of the column
- $V_t$  = Elution volume for very small molecules





# **COMPONENTS OF A SEC**

- 1. Stationary Phase**
- 2. The Mobile Phase**
- 3. The Columns**
- 4. The Pump**
- 5. Detectors**



# STATIONARY PHASE:

- Stationary Phase Semi-permeable, porous beads with well-defined range of pore sizes .
- Beads are crosslinked polymers
- Degree of crosslinking is controlled carefully to yield different pore sizes.
- Smaller pore sizes are used for rapid desalting of proteins or for protein purification.
- Intermediate pore sizes are used to separate relatively small proteins.

- Very large pore sizes are used for purification of biological complexes.
- Stationary phase used for gel exclusion chromatography include dextran (Sephadex™), polyacrylamide and dextran-polyacrylamide (Sephacryl™).
- Each is available with a variety of different ranges of pore size in the beads, permitting separation of macromolecules of different size



# **A good stationary phase should have following properties:**

- ❖ It should be chemically inert.
- ❖ It should be inexpensive.
- ❖ It should not react with component to be separated.
- ❖ It should not react with eluent.
- ❖ It should be colorless, uniform in size and shape.
- ❖ It should be mechanically stable.

- ❖ **Soft gel** e.g.- dextran(Sephadex), Polyacrylamide gels  
Separation of proteins.
- ❖ **Semi-rigid gel** e.g.- bio beads  
Separation of non-polar polymers in non-polar solvents.
- ❖ **Highly rigid** gels and glasses  
Separation of polar systems.

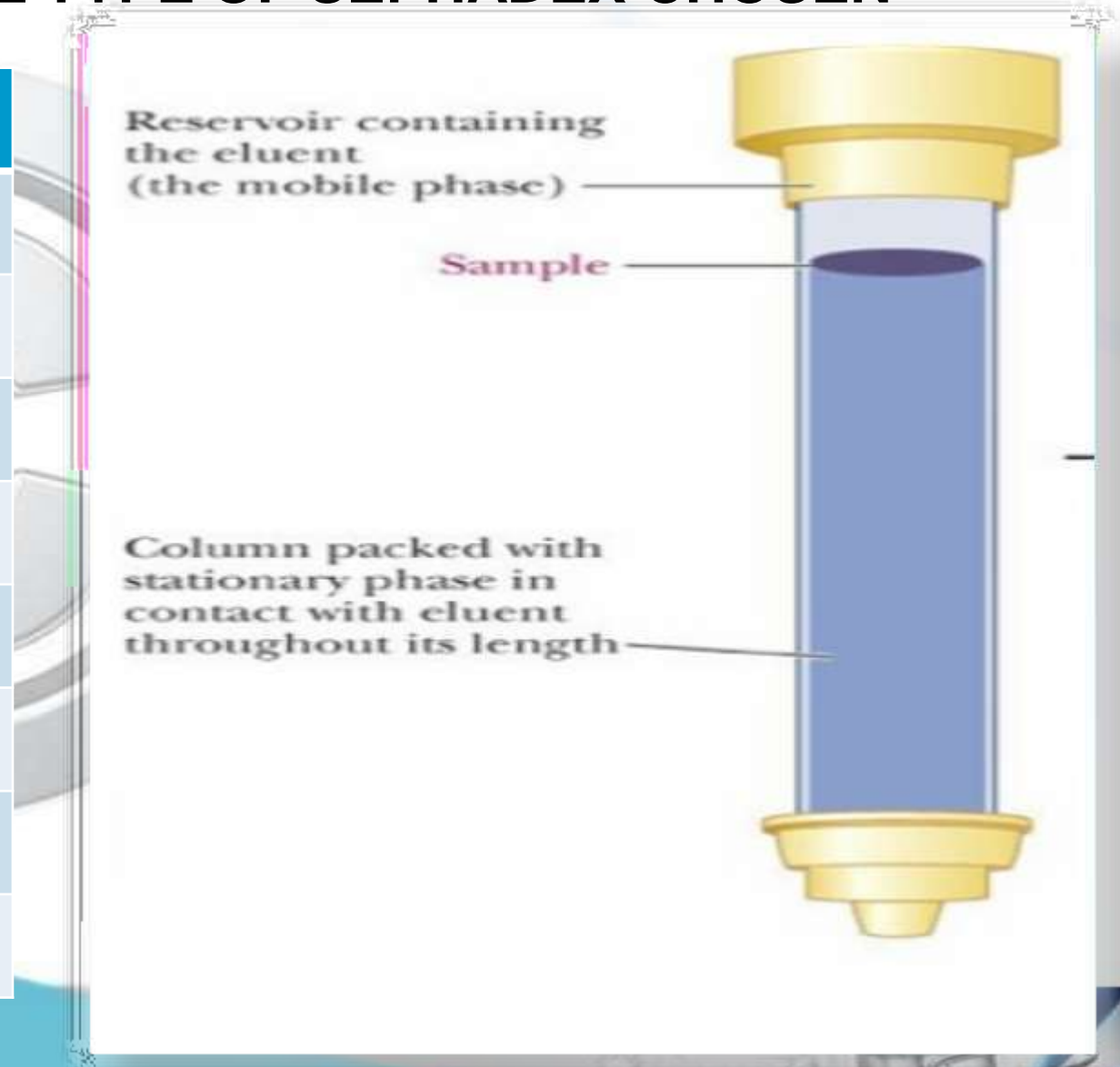
# Dextran

- A homopolysaccharide of glucose residues.
- It's prepared with various degrees of cross-linking to control pore size.
- It's bought as dry beads, the beads swell when water is added.
- The trade name is sephadex.
- It's mainly used for separation of small peptides and globular proteins with small to average molecular mass.



**TYPICAL SEPARATION RANGES THAT CAN BE ACHIEVED USING SEPHADEX ARE GIVEN BELOW. MOLECULES RANGING FROM 100 TO 600,000 DA CAN BE SEPARATED DEPENDING ON THE TYPE OF SEPHADEX CHOSEN**

SEPHADEX	RANGE
G10	100-800 Da
G15	(500-1500 Da),
G25	(1000-5,000 Da),
G50	(1,500-30,000 Da)
G75	(3,000-80,000 Da )
G100	(4,000-150,000 Da )
G150	(5,000-300,000 Da)
G200	(5,000-600,000 Da).



# Polyacrylamide

- these gels are prepared by cross linking acrylamide with N,N-methylene bis acrylamide.
- The pore size is determined by the degree of cross-linking.
- The separation properties of polyacrylamide gels are mainly the same as those of dextrans.
- They are sold as bio-gel P. They are available in wide range of pore sizes.



# Agarose

- Linear polymers of D-galactose and 3,6 anhydro-1-galactose.
- It forms a gel that's held together with H bonds. It's dissolved in boiling water and forms a gel when it's cold.
- The concentration of the material in the gel determines the pore size.
- The pores of agarose gel are much larger than those of sephadex or bio-gel p.
- It's useful for analysis or separation of large globular proteins or long linear molecules such as DNA



# Mobile phase

- The liquid used to dissolve the biomolecules to make the mobile phase is usually called a **buffer**.
- The mixture of biomolecules dissolved in the buffer is called the **sample**.
- The choice of mobile phase to be used in any separation will depend on the type of separation to be achieved and component to be separated.
- ❖ The most common eluents in for polymers that dissolve at room temperature.**e.g.**-Tetrahydrofuran,Chloroform, Dimethyl formamide.

# MOBILE PHASE

material	solvent
Synthetic elastomers (polybutadiene , polyisoprene )	Toluene
PS, PVC, Styrene-Butadiene Rubber , Epoxy resins	Tetrahydrofuran
Polyolefins	Tri- chloro -benzene
Polyurethane	Di- methylformamide
Proteins, polysaccharides	Water / Buffers

# SOLVENT SELECTION

The solvents used for mobile phase of SEC are limited to those follows following criteria:

- The solvent must dissolve the sample completely.
- The solvent has different properties with solute in the eluent: typically with solvent refractive index (RI) .
- solvent must not degrade the sample during use Otherwise, the viscosity of eluent will gradually increase over times.
- The solvent is not corrosive to any components of the equipment



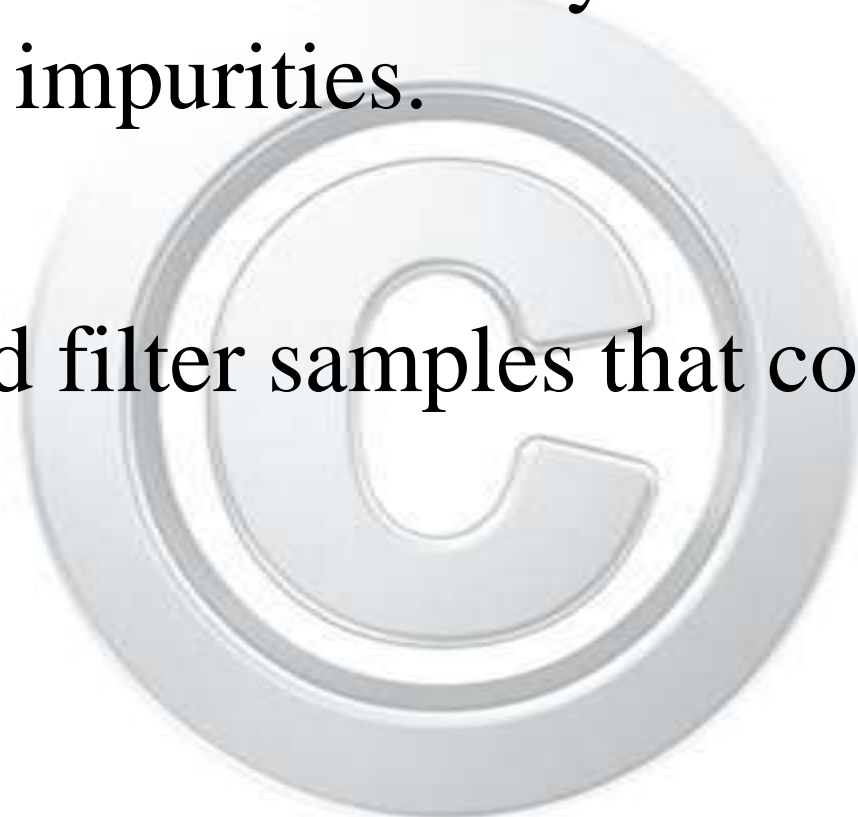
# Mobile Phase Preparation

- high purity of solvent is recommended.
  - Filter mobile phase solvents using 0.5 micron filter to remove any particular impurities such as dusts, insoluble salts.
  - Antioxidant is added to trichlorobenzene to keep solvent stable in high temperature.
  - Other additives eliminate adsorption or interaction of solutes with column packing materials
- Size Exclusion Chromatography

# **Sample preparation**

- The sample solutions are supposed to be prepared in dilute concentration (less than 2 mg/mL)
- A good solvent can dissolve a sample in any proportion in a range of temperatures.
- Samples with broad molecular weight distribution may require higher concentrations.
- It is recommended to filter the sample solutions before injecting into columns in order to get rid of clogging and excessively high pressure problems.

- Agitation and filtration Generally filtration is required to remove insoluble impurities.
- Do not agitate and filter samples that contain very high MW (>1 million).



# COLUMNS

## Commercially Available Columns

- ❖ analytical column- 7.5–8mm diameters.
- ❖ Preparative columns-22–25mm for.
- ❖ Usual column lengths-25, 30, 50, and 60 cm.
- ❖ Recently, narrow bore columns- 2–3mm diameter have been introduced, which save time and solve

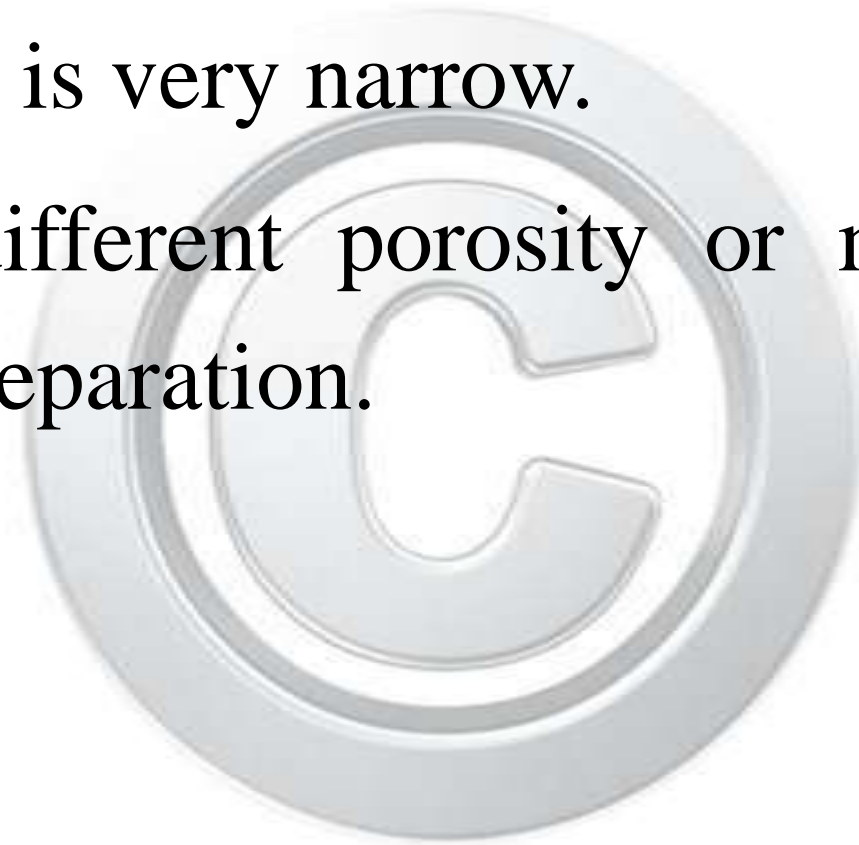




# **Selecting SEC column**

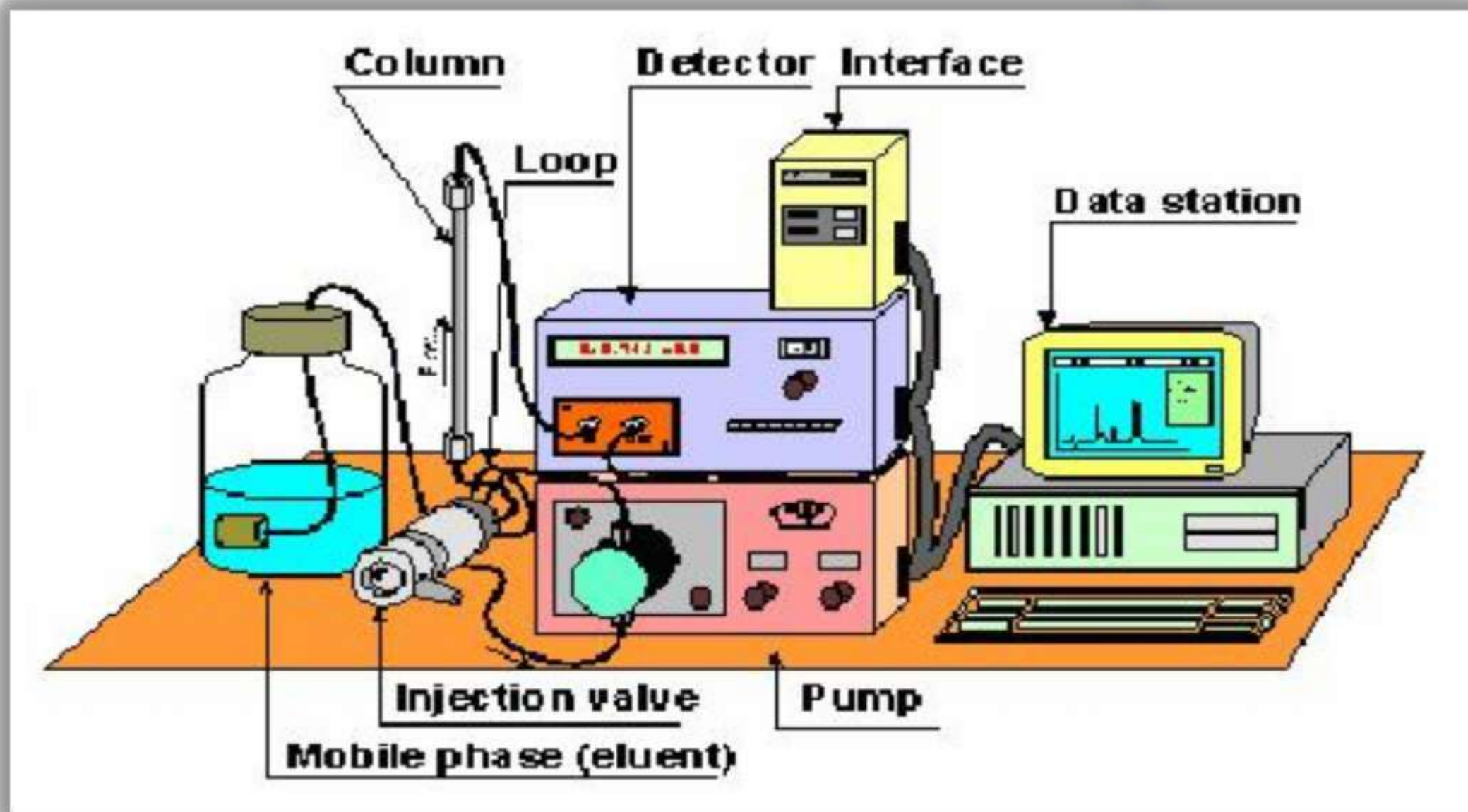
- ❖ Shorter columns save time and solvent.
- ❖ Small particles (typically 5 mm) provide a better resolution.
- ❖ On the other hand, 5 mm (or even 3 mm) packings are more sensitive towards contamination by samples containing impurities.
- ❖ Particles as large as 20 mm have been recommended for very high-molecular-weight polymers.
- ❖ ion.

- ❖ Small particle size packings can sometimes result in shear degradation of large polymer molecules because the space between particles is very narrow.
- ❖ Columns with different porosity or mixed-bed columns, provide a better separation.



# Handling SEC Columns

- ❖ A column set in SEC should be always run in the same mobile phase.(isocratic)
- ❖ SEC columns should never be operated in a backward direction.
- ❖ Care should also be taken in connecting columns or in sample injection.
- ❖ Replacing a clogged inlet frit is a dangerous operation which can reduce column performance.
- ❖ A damaged or dirty check valve of pump, can also reduce column life.





- ❖ A highly constant flow rate has to be maintained during the entire chromatogram. This is very important in SEC.
  - ❖ A change of the flow rate of only 0.1% can cause an error in molar mass of up to 10%.
  - ❖ Most pumps can only reproduce the flow rate to 0.2–0.3%.
  - ❖ In-line filters in the solvent reservoir may prevent particles from coming into the pump heads, which might damage the check valves or the pump seals.
- Types- **Syringe pumps, Reciprocating pumps**

# DETECTOR

## ❖ **Concentration sensitive detectors**

- Bulk Property Detectors- Refractive Index (RI) Detector
- Solute Property Detectors- Ultraviolet (UV) Absorption Detector
- Evaporative Detectors- Evaporative Light Scattering Detector (ELSD)

## ❖ **Molar mass sensitive detectors**

### ➤ Light Scattering Detectors

- Low Angle Light Scattering (LALS) Detectors
- Multiangle Light Scattering (MALS) detectors

➤ Viscosity Detectors- Differential Vscometers

**Other :-** Flame Ionization Detector (FID), A Mass Spectrometer or A Fourier Transform Infrared (FTIR) Spectromet



# ADVANTAGES

- ❖ Short analysis time.
- ❖ Well defined separation.
- ❖ Narrow bands and good sensitivity.
- ❖ There is no sample loss.
- ❖ Small amount of mobile phase required.
- ❖ The flow rate can be set.



# **DISADVANTAGES**

- ❖ Limited number of peaks that can be resolved within the short time scale of the GPC run.
- ❖ Filtrations must be performed before using the instrument to prevent dust and other particulates from ruining the columns and interfering with the detectors.
- ❖ The molecular masses of most of the chains will be too close for the GPC separation to show anything more than broad peaks
- ❖ .

# APPLICATION

- ❖ Proteins fractionation
- ❖ Purification
- ❖ Molecular weight determination.
- ❖ Separation of sugar, proteins, peptides, rubbers and others on the basis of their size.
- ❖ This technique can be determine the quaternary structure of purified proteins.



- ❖ SEC is a widely used technique for the purification and analysis of synthetic and biological polymers, such as protein, polysaccharides and nucleic acid.
- ❖ Various species of RNA and viruses have been purified using agarose gels.
- ❖ For Desalting
- ❖ For copolymerisation studies

